

146.3 °C (reported^{5b} mp 146–147 °C); [α]_D²⁰ -34.6° (c 0.940, CHCl₃).

Acknowledgments. We are very grateful to Professor T. Sakan, Institute of Food Chemistry, Osaka, Japan, and Professor A. Fujino, Osaka City University, who gave us a sample of the picrate of (-)-actinidine, and to Professor R. B. Woodward for his support during an early stage of this project.

Registry No.—1, 61899-98-7; 4, 564-24-9; 5, 61899-99-8; 7, 61900-00-3; 8, 7712-68-7; 9, 61900-01-4; 10, 61900-02-5; 12, 15524-81-9; 12 picrate, 61900-03-6; oxalyl chloride, 79-37-8.

References and Notes

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A Stereocontrolled Synthesis of (±)-Anhydronupharamine. The ¹H and ¹³C Nuclear Magnetic Resonance of Piperidine Nuphar Alkaloids¹

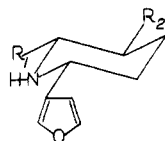
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Received December 27, 1976

(±)-Anhydronupharamine is prepared in 11 steps starting from 6-methyl-5-hepten-2-one and proceeding through key intermediates *trans*-3-methyl-2-(3-methyl-2-butenyl)cyclopentanone, *trans*-6-(3-methyl-2-butenyl)-5-methyl-2-piperidone, and *trans*-2-(3-methyl-2-butenyl)-3-methyl-6-(3-furyl)-2,3,4,5-tetrahydropyridine. Stereocontrol is based on the greater stability of *trans* substituents in a 2,3-disubstituted cyclopentanone and the more favorable reduction of a C-2 substituted 2,3,4,5-tetrahydropyridine from the direction opposite the C-2 substituent. The ¹H and ¹³C NMR characteristics of the various 3-furyl-substituted piperidines obtained in the course of synthesis are given and briefly discussed with regard to conformation.

The structures of (-)-anhydronupharamine (**1**) and (-)-nuphenine (**2**) exemplify the two stereochemical types of Nuphar piperidine alkaloids. The *trans* disposition of C-2 and C-3 hydrogen atoms in **1** similarly occurs in the Nuphar quinolizidine alkaloids where the carbons of the second ring might be considered constituted by those of the C-2 side chain in **1**. This *trans* arrangement appeared, until recently, to be the only one in the quinolizidine Nuphar alkaloids. However, the results of new isolation work show that the C-2 and C-3 *cis* arrangement of hydrogen atoms in **2** also presents itself in the C₁₅ quinolizidine 1-*epi*-deoxynupharidine² and in some C₃₀ thiaspiranes such as 1-*epi*,1'-*epi*-thiobinupharidine.³ Regardless of the steric disposition of the C-2 and C-3 substituents, the 3-furyl group at C-6 always assumes an equatorial conformation and is *cis* to the C-2 substituent in the naturally occurring Nuphar piperidines and quinolizidines.

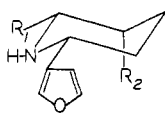


1, R₁ = (CH₃)₂C=CHCH₂; R₂ = CH₃

20, R₁ = R₂ = H

21, R₁ = CH₃; R₂ = H

28, R₁ = (CH₃)₂C(OH)CH₂CH₂; R₂ = CH₃



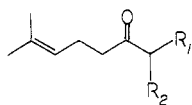
2, R₁ = (CH₃)₂C=CHCH₂; R₂ = CH₃

29, R₁ = (CH₃)₂C(OH)CH₂CH₂; R₂ = CH₃

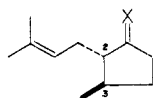
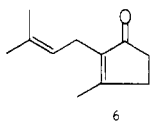
We sought to prepare the piperidine Nuphar alkaloids by routes which would offer control over the C-2, C-3, and C-6 stereochemistry and which appeared to hold some promise for appropriate elaboration of the C-2 side chain in order that the route could be extended later to the Nuphar quinolizidines. We report here the synthesis⁴ of (±)-anhydronupharamine by a route through which the stereocontrol of C-2 and C-3 substituents rests on the far greater stability of *trans* C-2, C-3 alkyl substituents in a cyclopentanone.⁵ As results were to demonstrate, the basis for the C-2, C-6 *cis* arrangement of substituents is the more favorable reduction of a C-2 substituted 2,3,4,5-tetrahydropyridine from the side opposite the C-2 substituent. In addition we report on the results of the ¹H and ¹³C NMR investigations of the stereochemistry of the new piperidine compounds which have arisen in the course of the synthesis.

Results and Discussion

Synthesis. The cyclopentanone **6**, substituted by γ,γ -dimethylallyl and methyl groups at C-2 and C-3, was prepared by starting from the 6-methyl-5-hepten-2-one (**3**) and proceeding through **4** and **5** according to an established sequence⁶ for preparing 2,3-disubstituted cyclopentanones. Thereafter the key intermediate cyclopentanone **7** possessing C-2 and C-3 *trans* substituents was prepared through lithium/liquid ammonia reduction of the cyclopentanone. None of the *cis* isomer



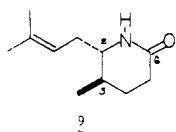
- 3, $R_1=R_2=H$
 4, $R_1=COOCH_3$; $R_2=H$
 5, $R_1=COOCH_3$; $R_2=CH_2COCH_3$



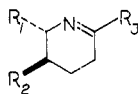
- 7, $X=O$
 8, $X=NOH$

could be detected by 1H NMR or GLC. The yields for these steps and all subsequent ones are given in the Experimental Section.

The conversion of ketone 7 to the oxime 8 with hydroxylamine hydrochloride in refluxing pyridine and subsequent Beckmann rearrangement of the oxime, by treatment of the latter with phosphorus pentachloride, achieved nitrogen incorporation in a six-membered lactam, 9, in the desired position relative to C-2 and C-3 as indicated by the 1H NMR. The trans disposition of substituents was largely preserved, but some loss of stereochemical integrity occurred in oxime formation. The presence of 8.5% of the cis oxime was detected by 1H NMR observation of the C-3 methyl doublet which appeared at δ 0.82 ppm, while the methyl doublet from the predominant trans isomer appeared slightly downfield at δ 1.00 ppm. This cis isomer was carried through the remainder of the synthesis.



The major obstacle to the completion of the synthesis was the attachment of a 3-furyl group to the carbon present as the lactam carbonyl carbon in 9. Alkylation of a lactam carbonyl carbon has been achieved through conversion of the lactam to a thioimide ester followed by treatment of the latter with a lithium alkyl in the presence of diisopropylaluminum hydride (DIBAH).⁷ Similarly the treatment of the *S*-methylthiolactim 10 with phenyllithium in the presence of DIBAH or diphenylmercury gave the imine 11 in yields up to 50%. The *O*-methylactim 12 and phenyllithium also produced 11 in the

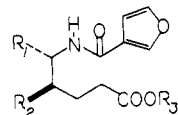


- 10, $R_1=R_2=H$; $R_3=SCH_3$
 11, $R_1=R_2=H$; $R_3=C_6H_5$
 12, $R_1=R_2=H$; $R_3=OCH_3$
 13, $R_1=R_2=H$; $R_3=3\text{-furyl}$
 14, $R_1=CH_3$; $R_2=H$; $R_3=OCH_3$
 19, $R_1=CH_3$; $R_2=H$; $R_3=3\text{-furyl}$
 23, $R_1=(CH_3)_2C=CHCH_2$; $R_2=CH_3$; $R_3=OCH_3$
 26, $R_1=(CH_3)_2C=CHCH_2$; $R_2=CH_3$; $R_3=3\text{-furyl}$

presence or absence of DIBAH, though in better yield when DIBAH was present.⁸ However when 3-furyllithium and DIBAH or bis(3-furyl)mercury replaced the phenyl counterparts, the corresponding 3-furyl-substituted imine 13 was not obtained.

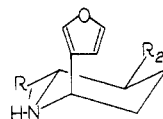
Since the direct attachment of the 3-furyl portion of the molecule seemed frustrated, the indirect incorporation of this group was attempted and achieved through *N*-acylation of an

O-methylactim with 3-furoyl chloride followed by decarboxylation and simultaneous nitrogen-carbon bond formation to reestablish the six-membered heterocyclic ring. Thus treatment of the *O*-methylactims 12 and 14 with lithium hydride and 3-furoyl chloride produced the amido esters 15 and 16, respectively,⁹ which after ester hydrolysis and pyrolysis of the resulting amidocarboxylic acids, 17 and 18, in the



- 15, $R_1=R_2=H$; $R_3=CH_3$
 16, $R_1=R_3=CH_3$; $R_2=H$
 17, $R_1=R_2=R_3=H$
 18, $R_1=CH_3$; $R_2=R_3=H$
 24, $R_1=(CH_3)_2C=CHCH_2$; $R_2=R_3=CH_3$
 25, $R_1=(CH_3)_2C=CHCH_2$; $R_2=CH_3$; $R_3=H$

presence of calcium oxide gave the imines 13 and 19 in 52 and 60% yields, respectively, in the last step. Sodium borohydride reduction of 13 gave 2-(3-furyl)piperidine, 20, while imine 19 led to a mixture of *cis*- (21, 91%) and *trans*-6-methyl-2-(3-furyl)piperidine (22, 9%). The sequence of transformations



- 22, $R_1=CH_3$; $R_2=H$
 27, $R_1=(CH_3)_2C=CHCH_2$; $R_2=CH_3$

was applied thereafter to the lactam 9. Conversion of the latter to the *O*-methylactim 23, *N*-acylation and ring opening to amido ester 24, hydrolysis to the amido acid 25, pyrolysis with calcium oxide to imine 26, and reduction with sodium borohydride produced a mixture of piperidines. According to GLC analysis, this mixture consisted of 85% anhydronupharamine (1) and 15% of the stereoisomers nuphenine (2) and the 2,6-*trans* isomer, 27. Each of the three was separated by elution chromatography and the samples of anhydronupharamine and nuphenine possessed chromatographic and spectrometric properties identical with those of the naturally occurring alkaloids.

1H and ^{13}C NMR. (–)-Anhydronupharamine has been correlated¹⁰ with (–)-nupharamine (28), which in turn has been correlated¹¹ with (–)-deoxynupharidine, whose stereochemistry is secure.¹² Therefore there is no question regarding the relative configuration of anhydronupharamine. The relative configurations of nuphenine¹³ and the closely related tertiary alcohol 3-*epi*-nupharamine¹⁴ (29) rest on the 1H NMR spectra, which indicate the C-3 methyl groups are axial. The same spectra are consistent with the C-2 side chains being equatorial, though the axial disposition of these side chains is not necessarily ruled out. We have found that a comparison of the 1H and ^{13}C NMR of the various 3-furyl-substituted piperidines has been useful in confirming and assigning conformation and configuration to the various 3-furyl-substituted piperidines.

The 1H doublets exhibited by the C-3 methyls in anhydronupharamine, 1, and the 2,6-*trans* isomer, 27, are at nearly the same field strength (δ 0.91 and 0.89 ppm, respectively) and have the same coupling constants (6.0 Hz), but are upfield from and have slightly smaller coupling constants than the δ 0.98 ppm methyl doublet ($J = 7.2$ Hz) shown by nuphenine 2. These 1H NMR properties of the C-3 methyl-substituted piperidines parallel those of the corresponding quinolizidines where axial methyl groups, substituted at C-1 or C-3 in chair

Table I. ¹³C Chemical Shift Values ^a of Selected Carbons in 3-Furyl-Substituted Piperidines

Compd	Substituted	Carbon no. ^b					CH ₂	CH ₃
		2 (6)	3 (5)	4	5 (3)	6 (2)		
	Piperidine ^c	47.7 (t)	27.5 (t)	26.1 (t)	27.5 (t)	47.7 (t)		
20	Mono	53.1 (d)	33.7 (t)	24.7 (t) ^d	25.9 (t) ^d	47.0 (t)		
21	Di	53.6 (d)	33.9 (t) ^e	24.9 (t)	33.2 (t) ^e	52.8 (d)		22.9
22	Di	46.9 (d)	31.0 (t)	20.0 (t)	33.0 (t)	45.5 (d)		21.3
1	Tri	53.8 (d)	34.7 (t) ^f	34.3 (t) ^f	35.9 (d)	64.1 (d)	32.4 (t)	18.2 ^g (18.5) (q) ^g
2	Tri	54.4 (d)	28.6 (t)	33.2 (t)	30.7 (d)	60.5 (d)	32.6 (t)	11.8 (q)
27	Tri	43.6 (d)	29.4 (t) ^h	29.2 (t) ^h	35.6 (d)	57.2 (d)	32.3 (t)	18.2 ⁱ (18.8) (q) ⁱ

^a Given in parts per million from δ 0.0 ppm from Me₄Si with multiplicity in parentheses. ^b The carbon to which the 3-furyl group is attached is C-2 in the mono- and disubstituted piperidines but C-6 in the trisubstituted piperidines. The remaining carbons in the ring are numbered in sequence accordingly. ^c Values taken from ref 16. ^{d-i} Assignments may be interchanged where the same superscript letter appears.

form rings, appear downfield with slightly larger coupling constants than equatorial methyls.¹⁵ Thus our analysis of the piperidines points to an equatorial C-3 methyl in anhydronupharamine and the 2,6-trans isomer, but an axial C-3 methyl in nuphenine.

The resonance of the proton attached to the 3-furyl bearing carbon appears in the region of δ 3.57–3.60 ppm as a doublet of doublets ($J = 8.0$ – 10.9 and 2.1 – 5.5 Hz) in the spectra of anhydronupharamine, nuphenine, and *cis*-2-(3-furyl)-6-methylpiperidine (21), while the corresponding resonance appears at lower field, δ 4.11 and 4.00, as a triplet ($J = 4.0$ Hz) in the spectra of the *trans*-2,6 compounds, 22 and 27. The higher field doublet of doublet resonance indicates an axial proton (3-furyl equatorial) split by vicinal axial and equatorial protons, but the lower field triplet indicates an equatorial proton (3-furyl axial) split by vicinal axial and equatorial protons having equal coupling constants.

The ¹H resonance of the second carbonyl proton adjacent to nitrogen is less straightforward in providing useful stereochemical information. This proton appears as a quintet of doublets at 2.78 and 3.04 ppm, respectively, in the spectra of the *cis*- and *trans*-2-(3-furyl)-6-methylpiperidines. The splitting pattern is best rationalized for both spectra by the proton in question being axial and split by each of the three methyl protons and the vicinal axial proton by the same amount, 6.4 Hz, and split again by the vicinal equatorial proton by 2.3 Hz. The lower field shift value of this proton in the 2,6-trans isomer 22 would seem to reflect the conformation of the 3-furyl group. In the nuphenine case, the proton appears as a triplet of doublets ($J = 7.4$ and 2.2 Hz) at δ 2.77 ppm. The splitting with the vicinal C-3 equatorial proton is ambiguous regarding the question whether the C-2 proton is axial or equatorial. However the chemical shift value of the proton in question agrees with that of the corresponding proton in the *cis*-2,6 model compound 21, and therefore suggests that the C-2 proton is axial and the side chain equatorial. In the case of anhydronupharamine and its *trans*-2,6 isomer 27, the chemical shift value of the C-2 proton is anomalously low, occurring coincidentally with the allyl methylene in the δ 2.0–2.6 ppm region. This anomalous chemical shift is occurring only when an equatorial methyl group is attached to C-3, but the nature of the influence which this group has on the C-2 proton is not clear.

The ¹³C chemical shifts, excluding the values for the 3-furyl, the vinyl, and the vinylmethyl carbons are given in Table I for the six 3-furyl-substituted piperidines. The chemical shifts excluded from Table I appear at the expected values.¹⁷ Assignments were made with the assistance of ¹H off-resonance decoupled spectra and the chemical shift comparison within the series. Assignments for the ring carbons of the *cis*- and *trans*-2,6 model compounds 21 and 22 were given additional support by the agreement of observed chemical shifts with

those calculated from parameters of Booth and Griffiths determined from a study of several methylpiperidines.¹⁸ Carbons adjacent to nitrogen in all compounds except the mono-substituted piperidine were distinguished by ¹H single frequency decoupling experiments.

A comparison of the ¹³C chemical shift values for the disubstituted piperidines 21 and 22 shows that all ring carbons, except C-5, are at higher field in 22 than in 21. In addition the C-6 methyl group chemical shift values are very nearly the same, although the one for 22 is slightly higher, by 1.6 ppm. These observations, along with the ¹H NMR splitting patterns and chemical shift values for C-6, are consistent with the predominant conformer of the *trans*-2,6 isomer 22 being the one possessing an axial 3-furyl group and an equatorial methyl group. The axial–equatorial chemical shift increment for a C-1 or C-3 methyl group in quinolizidines is about 6 ppm. Assuming that this increment can be applied to the 2-methylpiperidine case and using the 1.6-ppm value of the methyl chemical shift difference in 21 and 22, we estimate that the ratio of axial methyl conformer to axial 3-furyl conformer in the equilibrium mixture of the two is about 1:4.

The presence of the C-3 axial methyl in nuphenine is indicated by the lower chemical shift value of this group relative to the corresponding group in anhydronupharamine.¹⁹ Also consistent with this stereochemistry is the observation that all ring carbon chemical shift values, except that for C-6, are lower for nuphenine than anhydronupharamine. Similarly in comparing 27 with 1, both methyl chemical shift values are the same and the values for all ring carbons, except C-3, are lower in 27 than 1. These observations are consistent with the C-3 methyl group in 27 being equatorial and the C-6 3-furyl group being axial.

Experimental Section

Spectra were determined as follows: infrared (IR) neat on a Perkin-Elmer 137 spectrometer; ¹H NMR in CDCl₃ solution in 5-mm tubes (1% Me₄Si; δ 0.00) on Varian A60 A and XL 100-15 spectrometers, the latter operating at 100 MHz in the FT absorption mode, lock being established on CDCl₃ (m, s, d, t, q, q', and br refer to multiplet, singlet, doublet, triplet, quartet, quintet, and broad, respectively); ¹³C NMR on a XL 100-15 spectrometer operating in the FT absorption mode at 25.2 MHz employing 8192 data points, using 5–69-mg samples in 5-mm tubes, the CDCl₃ also furnishing the secondary reference signal (77.2 ppm from Me₄Si at δ 0.0 ppm) and the deuterium resonance for field-frequency lock. Fully ¹H noise decoupled, selective ¹H decoupled, and ¹H off-resonance decoupled ¹³C NMR spectra were obtained from 6 to 272 K transients, the number of transients for off-resonance decoupled spectra being at the higher end of the range. In all cases ¹³C spectral widths were 5000 Hz and acquisition times were 0.8 s. Pulse angles for ¹³C determinations ranged from 20 to 45 °C. Gas-liquid chromatography (GLC) is given as retention time (R_t) in minutes and was performed at the column temperature and flow rate or back pressure indicated on: a 5 ft \times ¼ in. stainless steel column packed with 1.5% OV101 on 100/120 Chromo-

sorb G HP (column A); a 5 ft \times 1/4 in. stainless steel column packed with 20% SE 30 on 60/80 Chromosorb W (column B); 5 ft \times 1/4 in. stainless steel column packed with 10% Carbowax 20M on Chromosorb G (column C); 5 ft \times 1/4 in. stainless steel packed with 2% OV101 on 100/120 Chromosorb G HP (column D); 10 ft \times 1/8 in. stainless steel column packed with 1.5% OV101 on 100/120 Chromosorb G (column E). Thin layer chromatography was performed on: A, Analtech pre-coated silica gel G (250- μ m thickness) developed with hexane-ether (4:1), or with the solvent system indicated, and visualized with I₂ vapor; B, Merck Alumina (250- μ m thickness) developed with hexane-ether (4:1) and visualized with Dragendorff-Munier reagent; C, Merck Alumina (250- μ m thickness) developed with 5% EtOAc in benzene and visualized with Dragendorff-Munier reagent. Elemental analyses were performed by Galbraith Analytical Laboratories, Knoxville, Tenn.; melting points (mp) were taken on a Mel-temp apparatus and are uncorrected.

Methyl 7-Methyl-3-oxo-6-octenoate (4). To a suspension of 38 g of NaH (1.58 mol) in 250 mL of dry ether under N₂ was added 142.75 g of dimethyl carbonate (1.58 mol), the resulting mixture was heated to reflux, and 100 g of 6-methyl-5-hepten-2-one (0.79 mol) (purchased from the Aldrich Chemical Co.) was added over 5 h to the heated mixture. The mixture was heated to reflux another 2 h, cooled to 25 °C, and poured onto crushed ice containing 99 mL of acetic acid. The ether solution was separated, washed with dilute aqueous NaHCO₃ and dried (MgSO₄). Removal of the ether at reduced pressure left an oil which was distilled to give 112 g of 4 (77%): bp 84–85 °C (0.5 mm); IR (neat) 1742 (COOCH₃), 1715 cm⁻¹ (RCOR); ¹H NMR δ 5.08 (m, 1 H, C-6 H), 3.70 (s, OCH₃), 3.41 (s, 2 H, C-2 H), 2.00–2.83 (m, 4 H, C-4 and C-5 H), 1.65 (d, *J* = 1.2 Hz, 3 H, CH₃), 1.60 (s, 3 H, CH₃); GLC (column A, 125 °C, back pressure 16 psi) *R*_t 6.9; TLC (A) *R*_f 0.39. Anal. Calcd for C₁₀H₁₆O₃: C, 65.19; H, 8.77. Found: C, 65.30; H, 8.87.

4-Carbomethoxy-9-methyl-8-decene-2,5-dione (5). To a stirred suspension of 5.72 g of NaH (0.237 mol) in 300 mL of ether under N₂ was added dropwise a solution of 44 g of 4 (0.237 mol) in 100 mL of ether. The resulting slurry was stirred until no effervescence was observed. Ether (200 mL) was added to facilitate stirring, the slurry was cooled to -25 °C, and a solution of 32.7 g of bromoacetone (0.238 mol) in 50 mL of ether was added with vigorous stirring. The mixture was heated to reflux for 30 min, cooled to 25 °C, and poured into crushed ice. The pH was adjusted to 5 with 25% aqueous H₂SO₄, the ether solution was separated, and the aqueous phase was extracted repeatedly with ether. The extracts were combined with the ether solution and the resulting solution was dried (MgSO₄). Removal of the ether at reduced pressure yielded 51 g of 5 (94%), 46 g of which was used in the next step without further purification and 1 g of which was distilled by short-path distillation to afford 5: bp 104–106 °C (0.1 mm); IR (neat) 1742 (COOCH₃), 1720 (RCOR), 1715 cm⁻¹ (RCOR); ¹H NMR δ 5.08 (m, 1 H, C-6 H), 4.00 (t, 1 H, C-2 H), 3.69 (s, 3 H, OCH₃), 3.50–2.00 (m, 6 H, C-4, C-5, and C-2 H), 2.14 (s, 3 H, CH₃), 1.65 (d, 3 H, CH₃), 1.60 (d, 3 H, CH₃); GLC (column B, 175 °C, 60 ml/min) *R*_t 9.4; GLC (column A, 150 °C, back pressure 18 psi) *R*_t 11.2; TLC (A) *R*_f 0.16. Anal. Calcd for C₁₃H₂₀O₄: C, 64.96; H, 8.40. Found: C, 65.05; H, 8.50.

3-Methyl-2-(3-methyl-2-butenyl)-2-cyclopentenone (6). A solution of 50 g of 5 in 800 mL of 3% aqueous NaOH was stirred at 70 \pm 2 °C for 3 h and thereafter cooled to 25 °C. The pH was adjusted to 4.0 with 25% aqueous H₂SO₄ and the liberated ketone was extracted repeatedly with 100 mL of ether. The combined extracts were washed with brine and dried (MgSO₄). Removal of the ether at reduced pressure and distillation of the residue gave 28 g of 6 (78%): bp 76–77 °C (0.2 mm); IR (neat) 1691 (RCOR), 1640 cm⁻¹ (C=C); ¹H NMR δ 5.05 (m, 1 H, C-7 H), 2.86 (d, 2 H, C-6 H), 2.44 (m, 4 H, C-4 and C-5 H), 2.03 (s, 3 H, CH₃), 1.66 (s, 6 H, C-9 and C-10 H); GLC (column B, 175 °C, 60 ml/min) *R*_t 3.9; GLC (column C, 210 °C, 60 ml/min) *R*_t 13.3; GLC (column A, 125 °C, back pressure 16 psi) *R*_t 9.3; TLC (A) *R*_f 0.19. Anal. Calcd for C₁₁H₁₆O: C, 80.42; H, 9.83. Found: C, 78.46; H, 9.92.

trans-3-Methyl-2-(3-methyl-2-butenyl)cyclopentanone (7). A solution of 5 g of 6 in 50 mL of ether was added to 400 mL of liquid NH₃ in a flask at -78 °C. Small pieces of lithium wire totaling 1.7 g (0.24 g atom) were added over a period of 3 min with vigorous stirring. The blue reaction mixture was stirred an additional 10 min, solid NH₄Cl was added, vigorous stirring was continued for 15 min, and to the resulting white slurry was added slowly 100 mL of water. The flask was removed from the cooling bath and kept at 35–40 °C until the bulk of the NH₃ had evaporated. The ether layer was separated, the aqueous layer was extracted repeatedly with ether, and the combined extract and original ether layer was dried (MgSO₄). Removal of the ether at reduced pressure gave an oil which was distilled to yield 3.7 g of 7 (73%): bp 83–84 °C (3.5 mm); IR (neat) 1740 cm⁻¹ (RCOR); ¹H

NMR δ 5.08 (m, 1 H, C-7 H), 1.25–2.50 (m, 8 H, (C)₃CH and (C)₂CH₂), 1.62 (d, 3 H, CH₃), 1.60 (s, 3 H, CH₃), 1.11 (d, 3 H, C-3 CH₃); GLC (column B, 170 °C, 60 ml/min) *R*_t 2.8; GLC (column A, 125 °C, back pressure 15 psi) *R*_t 5.0; TLC (A) *R*_f 0.52. Anal. Calcd for C₁₁H₁₈O: C, 79.44; H, 10.93. Found: C, 79.25; H, 10.84.

trans-3-Methyl-2-(3-methyl-2-butenyl)cyclopentanone Oxime (8). A solution of 2.5 g of hydroxylamine hydrochloride (0.035 mol) in 10 mL of 50% EtOH was added to a solution of 5 g of 7 in 10 mL of pyridine. The mixture was heated to reflux for 15 min, cooled to 25 °C, and the reaction flask was evacuated at 60 °C to form an oil which was treated with 50-mL portions of ether. The combined ether solution was dried (MgSO₄) and the ether was removed at reduced pressure to give an oil which when distilled afforded 5.3 g of 8 (98%): bp 90–91 °C (0.2 mm); IR (neat) 3300 (OH), 1670 cm⁻¹ (C=N); ¹H NMR δ 5.20 (m, 1 H, C-7 H); 1.16–3.00 (m, 8 H, (C)₃CH and (C)₂CH₂), 1.66 (s, 3 H, CH₃), 1.59 (s, 3 H, CH₃), 1.01 (d, 3 H, CH₃); GLC (column A, 150 °C, back pressure 17 psi) *R*_t 5.6; TLC (A) *R*_f 0.37. Anal. Calcd for C₁₁H₁₉NO: C, 72.86; H, 10.58; N, 7.72. Found: C, 72.92; H, 10.50; N, 7.73.

trans-6-(3-Methyl-2-butenyl)-5-methyl-2-piperidone (9). A 6.9-g quantity of PCl₅ (0.032 mol) was added to 4 g of 8 (0.022 mol) in ether and the resulting slurry was stirred at 25 °C for 20 h. The reaction flask was evacuated at 25 °C to obtain an oil which was dissolved in 150 mL of CH₂Cl₂ and the resulting solution was poured onto crushed ice containing 100 mL of 10 N NaOH. The mixture was stirred and additional NaOH was added to pH 10–12. The phases were separated and the aqueous phase was extracted repeatedly with CH₂Cl₂. The combined extracts were dried (MgSO₄) and the solvent was removed at reduced pressure to obtain an oil which was chromatographed on silica gel (grade 62) using hexane-ether-methanol (80:15:5) to obtain 2.6 g of 9 (65%): IR (neat) 1645 cm⁻¹ (amide); ¹H NMR δ 5.11 (m, 1 H, C-8 H), 3.75 (m, 1 H, C-6 H), 1.16–2.66 (m, 7 H, (C)₃CH and (C)₂CH₂), 1.68 (d, 3 H, CH₃), 1.63 (s, 3 H, CH₃), 1.03 (d, 3 H, CH₃); GLC (column A, 175 °C, back pressure, 18 psi) *R*_t 5.8; TLC (A, hexane-ether-methanol, 80:15:5) (A) *R*_f 0.20. Anal. Calcd for C₁₁H₁₉NO: C, 72.86; H, 10.58; N, 7.72. Found: C, 72.69; H, 10.64; N, 7.80.

trans-2-(3-Methyl-2-butenyl)-3-methyl-6-methoxy-2,3,4,5-tetrahydropyridine (23). To a mixture of 2 g of 9 (0.011 mol) and 0.344 g of dimethyl sulfate (0.0027 mol) at 80 °C was added dropwise over a period of 15 min 1.049 g (0.79 mL, 0.0083 mol) of dimethyl sulfate. The resulting mixture was kept at 80 °C 3 h, 2.5 mL of benzene was added, and then the mixture was cooled to 25 °C. A solution of 0.493 g of NaOH (0.012 mol) in 1 mL of water was slowly added with stirring; thereupon the temperature rose to 55 °C. The mixture was stirred at 55–60 °C for 15 min. After cooling, the two phases were separated. The aqueous phase was extracted repeatedly with benzene, the combined extracts were dried (MgSO₄), the benzene was distilled off at 760 mm, and the residue was distilled. The fraction collected at 45–46 °C (0.1 mm) consisted of 1.48 g of 23 (69%): IR (neat) 1680 (C=N), 1210 cm⁻¹ (OCH₃); ¹H NMR δ 5.27 (m, 1 H, C-8 H), 3.62 (s, 3 H, OCH₃), 2.83–3.57 (m, 1 H, C-6 H), 1.17–2.55 (m, 7 H, (C)₃CH and (C)₂CH₂), 1.72 (d, 3 H, CH₃), 1.67 (s, 3 H, CH₃), 1.00 (d, 3 H, CH₃); GLC (column A, 150 °C, back pressure 18 psi) *R*_t 3.4. Anal. Calcd for C₁₂H₂₁NO: C, 73.77; H, 10.85; N, 7.17. Found: C, 73.55; H, 10.93; N, 7.23.

Methyl erythro-5-(3-Furamido)-4,8-dimethyl-7-nonenoate (24). A solution of 1.0 g of 23 (0.0051 mol) in 12 mL of THF was added to a suspension of 0.035 g of LiH (0.005 mol) in 10 mL of THF under N₂. After the mixture had been heated to reflux for 3 h and cooled to 25 °C, a solution of 0.668 g of 3-furoyl chloride (0.0051 mol) in 12 mL of THF was added and the mixture was stirred for 7 days at 25 °C. A 12-mL quantity of 6 N aqueous HCl was added slowly with stirring. After continued stirring for 15 min, the mixture was extracted repeatedly with CH₂Cl₂. The combined extracts were dried (MgSO₄ and NaHCO₃) and the solvent was removed at reduced pressure to obtain an oil which was chromatographed on Alumina (activity 1) with ether and thereby was afforded 0.74 g of oily 24 (47%): ir (neat) 3300 (NH), 3120 (furan CH), 1735 (COOCH₃), 1630 (CONH), 871 cm⁻¹ (furan); ¹H NMR δ 7.88 (m, 1 H, 3-furyl α -H), 7.36 (m, 1 H, 3-furyl α -H), 6.58 (m, 1 H, 3-furyl β -H), 5.85 (d, 1 H, NH), 5.10 (m, 1 H, C-7 H), 3.96 (m, 1 H, C-5 H), 3.64 (s, 3 H, OCH₃), 1.00–2.60 (m, 7 H, (C)₃CH and (C)₂CH₂), 1.65 (s, 3 H, CH₃), 1.57 (s, 3 H, CH₃), 0.91 (d, 3 H, C-4 CH₃); GLC (column A, 200 °C, back pressure 20 psi) *R*_t 15.6; TLC (A, ether) *R*_f 0.73. Anal. Calcd for C₁₇H₂₅NO₄: C, 66.41; H, 8.21; N, 4.55. Found: C, 66.16; H, 8.08; N, 4.30.

erythro-5-(3-Furamido)-4,8-dimethyl-7-nonenoic Acid (25). A suspension of 0.75 g of 24 in 20 mL of 5% aqueous KOH was heated to reflux for 2 h. The mixture was cooled to 5 °C, the pH was adjusted to 2 with 6 N aqueous HCl, and the mixture was extracted with four 50-mL portions of methyl isobutyl ketone (MIBK). The combined

MIBK extracts were washed with 25 mL of brine, dried (MgSO₄), and the MIBK was removed at reduced pressure to afford an oily residue, which was dried over P₂O₅ at 60 °C (0.5 mm) to produce 0.70 g of **25** (98%): IR (neat) 3318 (NH), 3122 (furan CH), 1710 (COOH), 1630 (CONH), 872 cm⁻¹ (furan); ¹H NMR δ 7.88 (m, 1 H, 3-furyl α-H), 7.36 (m, 1 H, 3-furyl α-H), 6.58 (m, 1 H 3-furyl β-H), 5.85 (d, 1 H, NH), 5.10 (m, 1 H, C-7 H), 3.96 (m, 1 H, C-5 H), 1.00–2.60 (m, 7 H, (C)₃CH and (C)₂CH₂), 1.65 (s, 3 H, CH₃), 1.57 (s, 3 H, CH₃), 0.91 (d, 3 H, C-4 CH₃). Anal. Calcd for C₁₆H₂₃NO₄: C, 65.45; H, 7.91; N, 4.77. Found: C, 65.52; H, 7.97; N, 4.87.

trans-2-(3-Methyl-2-butenyl)-3-methyl-cis-6-(3-furyl)piperidine (1). A 0.5-g quantity of **25** was mixed thoroughly with 0.5 g of CaO and placed in a Pyrex tube fitted with a side arm which projected far into an Erlenmeyer flask cooled at 0 °C. The Pyrex tube was heated by the gentle application of a microburner flame until the enamine distilled. To the enamine in 20 mL of absolute ethanol was added 240 mg of NaBH₄ under N₂ and the resulting mixture was stirred 5 h at 25 °C. The solvent was removed at reduced pressure, the residue was dissolved in 25 mL of CH₂Cl₂, 5 mL of H₂O was added, the two phase system was stirred for 10 min, the phases separated, the aqueous extracted repeatedly with CH₂Cl₂, and all CH₂Cl₂ solutions combined and dried. Removal of the CH₂Cl₂ at reduced pressure afforded 225 mg of oil (57%) which according to GLC (column A, 150 °C, back pressure 17 psi) contained 85% **1** (R_t 13.2 min) and 15% of a mixture of what later proved to be **2** and **27** (R_t 14.8). A 140-mg portion of this oil was chromatographed on neutral Alumina (4% H₂O) in a 8 × 1 cm column packed in C₆H₆. Elution with the following solvents in the specified quantities produced the fractions and milligram amounts indicated in parentheses: 1 mL C₆H₆, A1 (6); 70 mL C₆H₆, A2 (123); 30 mL C₆H₆, A3 (8); 30 mL CH₂Cl₂-C₆H₆ (1:1), A4 (3). Fraction A1, a brown oil, was the only one which was Dragendorff inactive and was discarded. Fraction A2 was chromatographed on neutral Alumina (3% H₂O) by eluting with 5% ether-hexane in the following amounts, constituting the fractions indicated in parentheses: 50 mL (B1), 100 drops (B2), 36 mL (B3), 30 mL (B4), 20 mL (B5), 220 mL (B6). Fractions B7 and B8 were eluted with 60 mL of chloroform each. Combined fractions B1–B3 consisted of a total of 12 mg of UV active but Dragendorff inactive material and were discarded. Combined fractions B4–B6 consisted of a total of 79 mg of pure **1**. Fraction B7 consisted of a 2-mg mixture of **1** and **2** and B8 consisted of a 16-mg mixture of **1** and **27**. The GLC (column A, 150 °C, back pressure 17 psi), the TLC (B and C), IR (CCl₄), ¹H NMR, and ¹³C NMR of **1** were identical with those of (–)-anhydronupharine and are as follows: GLC R_t 13.2; TLC (B) R_f 0.80; TLC (C) R_f 0.54; IR (CCl₄) 2770 (Bohmann bands), 873 cm⁻¹ (3-furyl); ¹H NMR δ 7.27–7.35 (3-furyl α-H), 6.36 (apparent t, J = 1.4 Hz, 1 H, 3-furyl β-H), 5.12 (t, J = 6 Hz, 1 H, CH=C), 3.57 (dd, J = 10.0 and 2.1 Hz, 1 H, C-6 ax H), 1.96–2.52 (m, 3 H), 1.70 (s, CH₃ C(C)=C), 1.64 (s, CH₃ C(C)=C), 0.91 (d, J = 6.0 Hz, 3 H, C-3 CH₃); ¹³C NMR, see Table 1; GLC (column E, 185 °C) R_t 10.15.

cis-2-(3-Methyl-2-butenyl)-3-methyl-cis-6-(3-furyl)piperidine (2). Fraction B7 was chromatographed on neutral Alumina (4% H₂O) in a 0.6 × 24 cm column packed in hexane. Elution with 5% ether in hexane in 25-, 15-, and 40-mL volumes yielded fractions C1 (0 mg), C2 (1 mg), and C3 (0.5 mg). Fraction C2 contained pure **2**: TLC (B) R_f 0.82; TLC (C) R_f 0.50; GLC (column A, 150 °C, back pressure 17 psi) R_t 14.8; GLC (column E, 185 °C) R_t 11.04; ¹H NMR δ 7.28–7.36 (3-furyl α-H), 6.41 (apparent t, 1 H, 3-furyl β-H), 5.12 (t, J = 7 Hz, 1 H, C=CH), 3.60 (dd, J = 5.0 and 8.0 Hz, 1 H, C-6 H), 2.77 (td, J = 7.4 and 2.2 Hz, 1 H, C-2 H), 2.05 (br t, J = 7.4 Hz, 2 H, CH₂C=C), 1.63 (s, CH₃C(C)=C), 1.71 (d, J = 0.5 Hz, CH₃C(C)=C), 0.98 (d, J = 7.2 Hz, 3 H, C-3 CH₃), and whose TLC, GLC, and ¹H NMR were identical with those of naturally occurring (–)-nuphenine.

trans-2-(3-Methyl-2-butenyl)-3-methyl-trans-6-(3-furyl)piperidine (27). Fractions A3 and B8 were combined (24 mg) and chromatographed on neutral Alumina (4% H₂O) in a 1 × 10 cm column. Elution with 5% ether-hexane was carried out in the following volumes constituting the fractions indicated in parentheses: 100 mL (D1), 150 mL (D2), 100 mL (D3). Fraction D4 was eluted with 100 mL of CHCl₃. Fraction D2 contained 5 mg of pure **27**: TLC (B) R_f 0.39; GLC (column E, 185 °C) R_t 11.34; IR (CCl₄) 873 cm⁻¹ (3-furyl); ¹H NMR δ 7.22–7.46 (m, 2 H, 3-furyl α-H), 6.28 (m, 1 H, 3-furyl α-H), 5.12 (apparent t, J = 7.2 Hz, 1 H, CH=C), 4.11 (t, J = 4 Hz, 1 H, C-6 H), 1.72 (s, CH₃ C(C)=C), 1.66 (s, CH₃ C(C)=C), 0.89 (d, J = 6.0 Hz, 3 H, C-3 CH₃); ¹³C NMR, see Table I.

Nuphenine and (–)-Anhydronupharine. These two alkaloids had been isolated in these laboratories from *N. luteum* subsp. *variegatum* and *N. japonicum*, respectively, and were purified by elution chromatography on Alumina immediately before comparison with synthetic samples. Optical rotations, ¹H NMR, and IR of these sam-

ples agreed with those reported earlier for (–)-nuphenine (**2**) and (–)-anhydronupharine (**1**). The ¹³C NMR of these two compounds are reported in Table I.

5-(3-Furamido)pentanoic Acid (17). Procedure A. A suspension of 500 mg of methyl 5-(3-furamido)pentanoate, **15**,⁹ in 10 mL of 2.25% aqueous KOH was heated to reflux for 2 h, during which time solution resulted. The mixture was cooled to 25 °C over the course of 1 h, the pH was adjusted to 2.0 with 25% aqueous H₂SO₄, and the mixture was repeatedly extracted with 50-mL quantities of methyl isobutyl ketone. The combined extracts were dried (MgSO₄) and the solvent removed at reduced pressure to obtain an oil which was dissolved in 25 mL of ether. From this solution crystals were formed on inducement. Drying these crystals at 40 °C at reduced pressure gave 400 mg of **17** (85%); mp 118–119 °C.

Procedure B. To a solution of 5 g of 5-aminopentanoic acid (0.04 mol) in 150 mL of H₂O was added 250 mL of methyl isobutyl ketone (MIBK) and the pH of the mixture was adjusted to 8.5 with 10% aqueous KOH. A 6.68-g sample of 3-furoyl chloride (0.05 mol) in 75 mL of MIBK was added in one portion with vigorous stirring. As the pH dropped it was adjusted to 8.0–8.5 by dropwise addition of 10% aqueous KOH. The reaction mixture was stirred until the pH remained constant (about 1.5 h), at which point the pH was adjusted to 2.0 with 25% H₂SO₄ and two phases were separated. The aqueous phase was extracted repeatedly with MIBK and all the MIBK extracts and solutions were combined and dried (MgSO₄). Removal of the solvent at reduced pressure gave an oil which, in the manner described in procedure A above, was transformed to 8.7 g of crystalline **17** (97%); mp 118–119 °C; IR (KBr) 3340 (NH), 1700 (COOH), 1620 (CONH), 875 cm⁻¹ (3-furyl); ¹H NMR (Me₂SO-*d*₆) δ 8.15 (m, 2 H, 3-furyl α-H and COOH), 7.70 (m, 1 H, 3-furyl α-H), 6.85 (m, 1 H, 3-furyl β-H), 3.32 (m, 2 H, C-5 H), 2.25 (m, 2 H, C-2 H), 1.52 (m, 4 H, C-3 and C-4 H). Anal. Calcd for C₁₀H₁₃NO₄: C, 56.85; H, 6.22; N, 6.63. Found: C, 57.00; H, 6.46; N, 6.56.

2-(3-Furyl)piperidine (20). A 1.5-g sample of **17** was mixed thoroughly with 1.5 g of CaO and the mixture was heated as described above in the preparation of **1**. The resulting distillate was collected in 25 mL of 10% aqueous HCl, the pyrolysis tube was rinsed with CHCl₃, and the solid residue was extracted with CHCl₃. The aqueous HCl solution and the combined CHCl₃ solutions were added to a separatory funnel. After shaking, the CHCl₃ layer was discarded, the aqueous layer was cooled to 5 °C, mixed with ether, and basified to pH 10 with 20% NaOH. The aqueous phase was extracted repeatedly and all ether extracts were combined and dried (MgSO₄). Removal of the ether at reduced pressure gave 550 mg of brown oily **13** (52%); GLC (column A, 150 °C, back pressure 12 psi) R_t 3.1. A 500-mg portion of **13** in 5 mL of dry EtOH under N₂ was mixed with 250 mg of NaBH₄ and the resulting heterogeneous mixture was stirred at 25 °C for 5 h. The EtOH was removed at reduced pressure, the residue was suspended in ether, water was added to the suspension, and the layers were separated. The aqueous layer was extracted with ether and all ether extracts were combined and dried (MgSO₄). Removal of the ether at reduced pressure gave 432 mg of brown oily **20** (85%); GLC (column A, 150 °C, back pressure 12 psi) R_t 2.1; IR 2920, 2840 (CH), 870 cm⁻¹ (3-furyl); ¹H NMR δ 7.35 (m, 2 H, 3-furyl α-H), 6.45 (m, 1 H, 3-furyl β-H), 2.50–3.80 (m, C-2 and C-6 H), 1.75 (6 H, m, C-3, C-4, and C-5 H); ¹³C NMR is given in Table I. Anal. Calcd for C₉H₁₄NO: C, 57.60; H, 7.52; N, 7.37. Found: C, 57.49; H, 7.71; N, 7.37.

5-(3-Furamido)hexanoic Acid (18). A suspension of 300 mg of methyl 5-(3-furamido)hexanoate⁹ in 6 mL of 2.25% aqueous KOH was treated as **15** in the preparation of **17** (procedure A) described above. In that manner was obtained 242 mg of **18** (80%); mp 112–113 °C; IR (KBr) 3300 (NH), 1700 (COOH), 1630 (CONH), 875 cm⁻¹ (3-furyl); ¹H NMR (Me₂SO-*d*₆) δ 8.31 (s, 1 H, COOH), 7.83 (m, 2 H, 3-furyl α-H), 6.97 (m, 1 H, 3-furyl β-H), 3.8–4.3 (m, 1 H, C-5 H), 2.00–2.45 (m, 2 H, C-2 H). Anal. Calcd for C₁₁H₁₅NO₄: C, 56.85; H, 6.72; N, 6.21. Found: C, 58.52; H, 6.77; N, 6.47.

cis- and trans-6-Methyl-2-(3-furyl)piperidine (21 and 22). A mixture containing 440 mg of **18** and 440 mg of CaO was treated as described above in the preparation of **1**. In this manner was obtained 191 mg of a light brown oily **19** (60%); GLC (column A, 150 °C, back pressure 16 psi) R_t 3.1. A 185-mg portion of **19** in 3 mL of dry EtOH under N₂ was mixed with 185 mg of NaBH₄ and the resulting suspension was treated as described above in the preparation of **1**. Thereby was obtained 150 mg of a light brown oily mixture of **21** and **22** (80%), GLC (column A, 150 °C, back pressure 16 psi), R_t 2.4 (91% **21**) and 3.1 (9% **22**), which was chromatographed on a neutral alumina (activity 2) column (1 × 11 cm) eluted with CH₂Cl₂, in 20 (E1), 7 (E2) and 170 mL (E3) fractions and with CHCl₃ in 75 (E4) and 100 mL (E5) fractions. Fraction E3, 69 mg, contained **21**: GLC (column E, 150 °C) R_t 5.27; TLC (Alumina, twice developed, once with CH₂Cl₂ then

CHCl₃) *R*_f 0.38; IR (CCl₄) 2778 (Bohlmann bands), 873 cm⁻¹ (3-furyl); ¹H NMR δ 7.31 (m, 2 H, 3-furyl α-H), 6.36 (m, 1, 3-furyl β-H), 3.60 (dd, *J* = 10.4 and 3.0 Hz, 1 H, C-2 H), 2.78 (q'd, 1 H, C-6 H), 1.05 (d, *J*, 6.6 Hz, 3 H, C-2 CH₃); ¹³C NMR, see Table I; high-resolution MS, obsd/calcd mass (formula) 165.1151/165.1152 (C₁₀H₁₅NO).

Fraction E4 was chromatographed on a 1 × 11 cm column of neutral fractions and a 30-ml CHCl₃ fraction (F4). Fraction F4 was chromatographed on a 0.7 × 2-cm column of neutral Alumina (activity 2) with 150 mL of CH₂Cl₂ (G1) and two 75-mL portions of CHCl₃-CH₂Cl₂ (1:9) (G2 and G3). Fractions F3 (3 mg) and G3 (7 mg) were combined and consisted of pure 22: GLC (column E, 150 °C) *R*_t 6.06; TLC (Alumina, twice developed, once with CH₂Cl₂ and then CHCl₃) *R*_f 0.31; IR (CCl₄) 2718 (very weak Bohlmann bands), 873 cm⁻¹ (3-furyl); ¹H NMR δ 7.24-7.44 (m, 2 H, 3-furyl α-H), 6.36 (m, 1 H, 3-furyl β-H), 4.10 (t, *J* = 4 Hz, 1 H, C-2 H), 3.04 (q'd, *J* = 7.0 and 2.2 Hz, 1 H, C-6 H), 1.11 (d, *J* = 6.0 Hz, 3 H, C-6 CH₃); ¹³C NMR, see Table I; high-resolution MS, obsd/calcd mass (formula) 165.1173/165.1152 (C₁₀H₁₅NO).

Registry No.—1, 61949-86-8; 2, 61949-87-9; 3, 110-93-0; 4, 53067-23-5; 5, 61900-43-4; 6, 61900-44-5; 7, 61900-45-6; 8, 61900-46-7; 9, 61900-47-8; 13, 61900-48-9; 15, 61586-90-1; 17, 61900-30-9; 18, 61900-31-0; 19, 61900-32-1; 20, 61900-33-2; 21, 61900-34-3; 22, 61900-35-4; 23, 61900-36-5; 24, 61900-37-6; 25, 61900-38-7; 27, 61949-85-7; dimethyl carbonate, 616-38-6; hydroxylamine HCl, 5470-11-1; dimethyl sulfate, 77-78-1; 3-furoyl chloride, 26214-65-3; 5-aminopentanoic acid, 660-88-8; 5-(3-furamido)hexanoate, 61900-39-8; (-)-nuphenine, 4850-01-5; (-)-anhydronupharine, 4849-88-1.

References and Notes

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A Carbon-13 Nuclear Magnetic Resonance Study of Thiol Esters

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Received December 14, 1976

The ¹³C NMR chemical shifts for each of the carbons of a number of simple thiol esters have been measured in Me₂SO-*d*₆ and CDCl₃ and the results related to known chemical properties of the thiol ester function. In general the chemical shift of the thiol ester carbonyl carbon occurs some 15–20 ppm further downfield than that found for all other carboxylic acid derivatives reported to date. The carbon α to the carbonyl function in thiol esters is also shifted downfield by about 10 ppm relative to the α carbon in acids, oxygen esters, or amides. The effect of carbon group and halogen substituents on thiol ester chemical shifts has been analyzed. A solvent study on β-hydroxy thiol esters shows that the carbonyl carbon is shielded in Me₂SO-*d*₆ relative to CDCl₃, which may be attributed to intramolecular hydrogen bonding in the latter solvent. Carbon-13 chemical shift changes resulting from conversion of mercaptans to thiol ester derivatives indicate relatively little difference between *S*-*tert*-butyl and other types of *S*-alkyl thiol esters in contrast to results obtained previously with *tert*-butyl oxygen esters.

The thiol ester group 1 is the ester function of choice in condensation and acyl transfer reactions occurring in biochemical systems.¹ In contrast to (oxygen) esters or amides, relatively little is known about the electronic structure of this group. As a result of our interest in the chemistry and properties of thiol esters, we have undertaken a ¹³C NMR study of this class of compounds. A search of the literature has not produced any general ¹³C NMR studies on the thiol ester

function.² We have thus obtained natural abundance ¹³C NMR spectra for some 30 different compounds. These results are discussed in connection with ¹³C NMR data available for other carbonyl derivatives.^{3,4} Substituent effects of the thiol ester group are analyzed and the effect of structure on thiol ester chemical shifts has been examined. Finally, we have focused attention on the relationship of these ¹³C NMR results to the chemistry and biological properties of the thiol ester function.

Experimental Section

Spectra. The ¹³C NMR spectra were obtained on ca. 20–25% (w/v) solutions in DCCl₃ or Me₂SO-*d*₆ using a Varian CFT-20 spectrometer

